Directions for Laboratory Reporting

The disease, events, and conditions reportable to Tennessee Department of Health (TDH) by laboratories for 2020, including laboratories in healthcare facilities, are provided in the 2020 List of Reportable Diseases in Tennessee: For Laboratories (Page 2 of the List). This Detailed Laboratory Guidance document (referenced in the List) provides additional details regarding the reportable tests and results, specimen source, and specimen/isolate submission to the Tennessee Department of Health Laboratory. Refer to https://www.tn.gov/health/health-program-areas/lab.html for additional details about submission. Please contact the TDH Laboratory at (615) 262-6300 before submitting specimens or isolates for pathogens requiring immediate notification. Please refer to Page 6 of this document for blood lead test reporting.

Laboratories should report via electronic laboratory reporting, a printed laboratory report, or online reporting.

- Requirements for electronic laboratory reporting are available at https://www.tn.gov/health/cedep/laboratory-reporting.html.

- Requirements for printed laboratory reports are below:

(1) Patient demographics (including patient date of birth, address, sex, race, ethnicity, and telephone)
(2) Ordering provider and facility name, phone number, address
(3) Performing laboratory name, phone number, and address
(4) Reporting facility name, phone number, address
(5) Date of the laboratory report
(6) Test performed (may differ from the test ordered)
(7) Accession number
(8) Specimen type/source and collection date
(9) Result (quantitative and qualitative), interpretation, and reference range

  - If the printed laboratory report does not include the required information listed above, then the PH-1600 should be completed for the missing information and submitted with the printed laboratory report.
  - Laboratories are only required to report specimen collection date and specimen source in the Clinical Information section.
  - The PH-1600 is available on the Reportable Diseases website at https://www.tn.gov/content/dam/tn/health/documents/reportable-diseases/PH-1600.pdf.
  - Laboratory reports and the PH-1600, if necessary, may be faxed directly to the local or regional health office (see https://www.tn.gov/health/health-program-areas/localdepartments.html) or the Communicable and Environmental Diseases and Emergency Preparedness (CEDEP) Division at (615) 741-3857.

- Online reporting is available via Morbidity Reports: https://hssi.tn.gov/auth/Login.
  - To sign up to use Morbidity Reports, please visit https://redcap.health.tn.gov/redcap/surveys/?s=8L7CMWHN4M.

For questions, contact CEDEP at (615) 741-7247 or (800) 404-3006.

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
Detection in one or more specimens of etiological agents of disease or conditions, not limited to those listed in this document, are of urgent public health significance!	Detection in one or more specimens of etiological agents of disease or conditions not limited to those listed in this document that are of urgent public health significance. Laboratories reporting outbreak events or conditions not listed in this Table but of public health significance should immediately contact the TDH Communicable and Environmental Disease Services via telephone at (615) 741-7247 or 1-800-404-3006.	By Request	L & P
Pan-nonsusceptible organisms and other unusual resistance !	Isolation from any specimen source that is pan-nonsusceptible or exhibits unusual resistance, regardless of active infection. Pan-nonsusceptible organisms are defined as an organism with susceptibility results from a validated instrument (or disc diffusion/ e-tests) that show resistant or intermediate interpretations to all antibiotics tested. Colistin and/or tigecycline susceptibility interpretation should not be considered when evaluating resistance. Examples of unusual resistance are shown in CLSI M100, Appendix A, Category 1. Isolates should be submitted urgently to the Tennessee Department of Health Laboratory. Please include identification and susceptibility test results (including numeric MIC values) with isolate. Contact hai.health@tn.gov for clarification/questions. See Appendix A of the M100 Performance Standards for Antimicrobial Susceptibility Testing.	Required	L & P
Acinetobacter species, Carbapenem-resistant **eip**	Acinetobacter species from any clinical specimen (including nonsterile sites and rectal/perirectal swabs) and non-susceptible isolates (intermediate or resistant to at least one carbapenem or PCR detection of carbapenemase-producing gene). Please include susceptibility test results (including numeric MIC values). Report only for residents of Davidson, Cheatham, Robertson, Sumner, Wilson, Rutherford, Dickson, or Williamson counties. Contact hai.health@tn.gov for clarification/questions. Isolates are required to be submitted to the Tennessee Department of Health Laboratory.	Required	L
Anaplasma phagocytophilum, species	Positive by any method for any specimen. Include speciation results if known.	--	L & P
Babesia species	Positive by any method for any specimen.	--	L & P
Bacillus anthracis !	Positive by any method (culture, IHC, serology, PCR, mass spectrometry, or other test) for any specimen for *Bacillus anthracis* or *Bacillus cereus* expressing anthrax toxins.	Required	L & P
Bordetella pertussis ☎	Positive culture or detected by nucleic acid amplification or polymerase chain reaction (PCR) for any specimen.	--	L & P
Borrelia burgdorferi	1) A positive culture for *B. burgdorferi* 2) A positive two-tier test. This is defined as a positive or equivocal enzyme immunoassay (EIA) or immunofluorescent assay (IFA) followed by a positive IgM or IgG Western immunoblot (WB) for Lyme disease. ▪ An IgM WB is considered positive when at least two of the following three bands are present: 24 kDa (OspC)*, 39 kDa (BmpA), and 41 kDa (Fla). Disregard IgM results for specimens collected >30 days after symptom onset. ▪ An IgG WB is considered positive when at least five of the following 10 bands are present: 18 kDa, 24 kDa (OspC)*, 28 kDa, 30 kDa, 39 kDa (BmpA), 41 kDa (Fla), 45 kDa, 58 kDa (not GroEL), 66 kDa, and 93 kDa. 3) A positive single-tier IgG WB test for Lyme disease (see above for how to identify a positive IgG WB). While a single IgG WB is adequate for surveillance purposes, a two-tier test is still recommended for patient diagnosis. *Depending upon the assay, OspC could be indicated by a band of 21, 22, 23, 24 or 25 kDA.	--	L & P
Brucella species ☎	Positive by any method (including culture, serology or others) for any specimen. Include quantitative antibody titer result indicating a positive test result when available. Isolates/specimens are required for submission to the Tennessee Department of Health Laboratory.	Required	L & P
Burkholderia mallei / Burkholderia pseudomallei !	Positive cultures for *Burkholderia mallei* and *Burkholderia pseudomallei* are reportable by laboratories. Isolates are required to be submitted to the Tennessee Department of Health Laboratory.	Required	L

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
California/LaCrosse serogroup viruses: California Encephalitis Virus, LaCrosse Encephalitis Virus, Jamestown Canyon Virus, Keystone Virus, Snowshoe Hare Virus, Trivittatus Virus	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus-specific neutralizing antibodies. Any specimen.	--	L & P
Campylobacter species	Positive by any method (including culture, EIA, & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Candida auris (including rule-out *Candida auris*) ☎	Positive by any method for any specimen. Detection from any site/specimens (including swabs from skin). Please note: *C. auris* can be misidentified as a number of different organisms when using traditional biochemical methods for yeast identification such as VITEK 2 YST, API 20C, BD Phoenix yeast identification system, and MicroScan. See https://www.cdc.gov/fungal/candida-auris/recommendations.html for greater detail. If species identity cannot be determined or one of the species shown in the table at above URL is identified, please contact HAI team at (615) 741-7247. Such isolates are considered "rule out *C.auris*" isolates. *C. auris* and "rule out *C.auris"* isolates should be submitted urgently to the Tennessee Department of Health Laboratory. Contact hai.health@tn.gov for clarification/questions.	Required	L & P
Chikungunya virus ☎	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus-specific neutralizing antibodies. Any specimen.	--	L & P
Chlamydia trachomatis	Positive by any method for any specimen.	--	L & P
Clostridium botulinum or botulinum toxin: Foodborne ! or Wound !	Positive by any method for any specimen.	Required	L & P
Clostridium botulinum or botulinum toxin: Infant	Positive by any method for any specimen.	Required	L & P
Clostridium difficile **eip**	Positive by any method for any specimen. Include methodology. Report only for residents of Davidson County. Contact hai.health@tn.gov for clarification/questions.	Requested	L & P
Clostridium tetani	Positive by any method for any specimen.	Required	L & P
Colistin-resistant (plasmid mediated) gram negative bacteria ☎	Positive by any method for any known plasmid-mediated colistin resistance mechanisms (e.g., *mcr-1* , *mcr-2*). Isolates/specimens from any specimen and body site (including screening tests to determine colonization). Excludes *Proteus, Providencia, Morganella,* and *Serratia* species. Submit isolates with MIC ≥4 for colistin.	Required	L & P
Corynebacterium diphtheria ☎ or *Corynebacterium ulcerans* ☎	Positive culture from any clinical source that has a positive Elek test for toxin production	Required	L & P
Coxiella burnetii ☎	Positive by any method (culture, IHC, serology, PCR, or other test) for any specimen is reportable. Include quantitative antibody titer result indicating a positive test result when available. Isolates are required to be submitted to the Tennessee Department of Health.	Required	L & P
Cryptosporidium species	Positive by any method for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Cyclospora species	Positive by any method **(including PCR)** for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
Dengue virus	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus- specific neutralizing antibodies. Any specimen.	--	L & P
Ehrlichia species (including *E. chaffeensis* and *E. ewingii*)	Positive by any method for any specimen. Include speciation results if known.	--	L & P
Enterobacteriaceae, Carbapenem-resistant	Any organism from the *Enterobacteriaceae* family (including but not limited to *Escherichia coli, Enterobacter* species, and *Klebsiella* species), from any clinical specimen (including rectal/perirectal swabs), resistant to at least one carbapenem antibiotic according to break points effective as of 2012 CLSI guidelines (i.e., ertapenem MIC ≥2.0 or doripenem/imipenem/meropenem MIC ≥ 4.0). Include all susceptibility results with the numeric MIC values (interpretation alone is insufficient), and all carbapenemase-production (mCIM, Carba NP or modified-Hodge) or resistance mechanism testing results (positive or negative). For example, polymerase chain reaction [PCR] or metallo-β-lactamase for Klebsiella pneumonia carbapenemase [KPC], New Delhi metallo-β-lactamase [NDM], Verona integron encoded metallo-β-lactamase [VIM], the imipenemase [IMP] metallo-β-lactamase, or OXA-48 carbapenemase). For *Proteus* spp., *Providencia* spp., and *Morganella morganii*: only submit isolates from these genera if elevated MICs are observed for ertapenem, meropenem, or doripenem, as these isolates exhibit intrinsic resistance to imipenem. Contact hai.health@tn.gov for clarification/questions.	Required	L & P
Enterococcus species, Vancomycin-resistant	Isolation of enterococci from any clinical specimen from a sterile site AND "Nonsusceptible" isolate (i.e., intermediate- or high level resistant) to vancomycin. Please include susceptibility test results with the numeric MIC values.	--	L & P
Equine encephalitis viruses: Eastern ☎, Venezuelan ☎, Western	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus- specific neutralizing antibodies. Any specimen.	--	L & P
Escherichia coli, Extended Spectrum Beta Lactamase-producing **eip**	*E. coli* resistant to at least one extended-spectrum cephalosporin (ceftazidime, cefotaxime or ceftriaxone), using the 2017 CLSI breakpoints. Include all susceptibility results with the numeric minimum inhibitory concentration values, and the results (positive or negative) of all carbapenemase-production or resistance mechanism testing. Report only for residents of the following counties: Lewis, Marshall, Maury, Wayne. Any specimen. Results should not be sent via electronic laboratory reporting. Contact hai.health@tn.gov for clarification/questions.	Requested	L
Escherichia coli, Shiga toxin-producing	Positive by any method (including culture, EIA, & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. For state public health labs, please include negative, not isolated, and no growth results. For any Shiga toxin-producing Escherichia coli (STEC), including *E. coli* O157s and *E. coli* non-O157s, EIA positive broths for shiga-like toxin will also be accepted. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Francisella tularensis ☎	Positive by any method for any specimen (including culture, fluorescent assay, PCR or serology) is reportable. Isolates are required to be submitted to the Tennessee Department of Health Laboratory.	Required	L & P
Haemophilus influenza ☎	Positive culture or PCR from a sterile site.	Required	L & P
Hepatitis A virus ☎	Positive IgM anti-HAV for any specimen. Nucleic Acid test (NAT) for hepatitis A virus RNA positive (including genotype testing) for any specimen. Include associated results for additional serological markers for hepatitis (including hepatitis B and C), alanine aminotransferase (ALT) and bilirubin levels if available.	--	L & P

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
Hepatitis B virus	• Positive hepatitis B surface antigen (HBsAg) • Positive IgM antibody to hepatitis B core antigen (IgM anti-HBc) • Positive hepatitis B "e" antigen (HBeAg) • Positive nucleic acid test for hepatitis B DNA (including qualitative, quantitative or genotype testing) And, if any of above tests are positive, also report: • Pregnancy status • All associated results (positive or negative) for additional serological markers of hepatitis (including hepatitis A and C), alanine aminotransferase (ALT) and bilirubin levels	--	L & P
Hepatitis C virus	• Positive or negative confirmatory assays (e.g. antigen or nucleic acid amplification testing for hepatitis C RNA [qualitative, quantitative or genotype testing]) • Positive hepatitis C antibody (anti-HCV) And, if any of above tests are positive, also report: • Pregnancy status • All associated results (positive or negative) for additional serological markers of hepatitis (including hepatitis A and B), alanine aminotransferase (ALT) and bilirubin levels	--	L & P
Human Immunodeficiency Virus (HIV)	o Reactive/positive/detectable tests for HIV - REPORT THE FOLLOWING - Screening test* and positive supplemental test** - p24 Antigen, nucleic acid detection (NAT/NAAT), viral culture, or genotype nucleotide sequences+ o CD4 count and percent - REPORT ALL RESULTS o HIV RNA (viral load) including undetectable (or equivalent)/detectable interpretation, quantitative count, and log count - REPORT ALL RESULTS If reporting any of the above, also report pregnancy status if available *Screening tests can include antibody (Ab) or antigen/antibody (Ag/Ab). **Supplemental tests can include western blot, immunofluorescent assay (IFA), conventional or rapid immunoassay (IA), or HIV-1/2 type-differentiating immunoassay (IA). If screening testing results are not available, report supplemental results. +Genotype nucleotide sequences are reportable by laboratories with ability to submit electronic lab reports (ELR). In accordance with T.C.A. §37-1-403, any physician or other person diagnosing or treating venereal herpes or any of these reportable sexually transmitted diseases in a child 13 years of age or younger should make a confidential written report of the case to the Department.	--	L & P
Influenza virus, detection of a novel or pandemic influenza A virus strain from a human **!**	Positive viral culture or PCR for any specimen.	Required	L & P
Klebsiella species, Extended Spectrum Beta Lactamase-producing **eip**	*Klebsiella* species, resistant to at least one extended-spectrum cephalosporin (ceftazidime, cefotaxime or ceftriaxone), using the 2017 CLSI breakpoints. Include all susceptibility results with the numeric minimum inhibitory concentration values, and the results (positive or negative) of all carbapenemase-production or resistance mechanism testing. Report only for residents of the following counties: Lewis, Marshall, Maury, Wayne. Any specimen. Results should not be sent via electronic laboratory reporting. Contact hai.health@tn.gov for clarification/questions.	Requested	L

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
Lead levels	All laboratories that run blood lead tests and those who conduct on site blood lead analysis with portable devices are to report all blood lead test results for Tennessee residents. Elevated blood lead levels (≥5 µg/dL) should be reported within 1 week and those <5 µg/dL should be reported within 1 month. Reports should include Patient's First Name, Last Name, Date of Birth, Gender, Race, Ethnicity, Address (Street Address, City, State, Zip Code and County of Residence), Sample Date, Sample Type, Result, Provider's Name and Phone Number and Payment Source. All blood lead test results may be reported electronically or via fax. For more information, refer to https://www.tn.gov/health/health-program-areas/mch-lead/for-providers.html or email UT Extension at leadtrk@utk.edu for assistance.	--	L & P
Legionella species	Positive by any method (culture, urine antigen, PCR, DFA, IHC, or serology) for any specimen is reportable. Isolates/positive specimens (expectorated sputum and bronchial lavage fluids only) are required for submission to the Tennessee Department of Health Laboratory.	Required	L & P
Listeria species	Positive by any method (including culture & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Measles virus !	Positive IgM or rising IgG titer or detected by nucleic acid amplification or positive viral culture for any specimen.	Required	L & P
Meningitis, isolation or demonstration of any bacterial species from cerebrospinal fluid ☎	Isolation of any bacteria from cerebrospinal fluid by culture, antigen, or PCR testing.	Required	L & P
Middle East Respiratory Syndrome Coronavirus (MERS- CoV) !	Positive by any method for any specimen.	Required	L & P
Mumps virus ☎	Positive IgM or rising IgG titer or detected by nucleic acid amplification or positive viral culture for any specimen.	Required	L & P
Mycobacterium leprae	Demonstration of acid-fast bacilli in skin or dermal nerve (using Fite stain) or identification of non-caseating granulomas with peripheral nerve involvement.	Required	L & P
Mycobacterium species other than *M. tuberculosis* (non-pulmonary sites only)	Any AFB smear, culture, HPLC, DNA probe or nucleic acid amplification test (NAAT) from any non-pulmonary site indicating presence of acid-fast bacilli. All specimens, except respiratory.	Requested	L & P
Mycobacterium tuberculosis complex (M. tuberculosis, M. bovis, M. africanum, M. canettii, M. microti)	The first AFB-positive respiratory specimen (by fluorochrome or acid-fast stain) indicating presence of acid-fast bacilli; submit specimen within 3 business days of collection.	Required	L & P
	Any specimen, from any site, with a positive nucleic acid amplification test (NAAT --- including, but not limited to: PCR, MTD, GeneXpert, MTBDR Plus [HAIN test]) indicating detection of *Mycobacterium tuberculosis* complex or associated point mutation from any site; submit specimen within 3 business days of test result. ☎	Required	L & P
	Any culture result by HPLC or DNA probe positive for *Mycobacterium tuberculosis* complex from any site; submit isolate within 5 business days of test result.☎	Required	L & P
	All anti-TB drug susceptibility results, by molecular or dilutional method, from a specimen or isolate from any site, with confirmed presence of *Mycobacterium tuberculosis*; anti-TB drugs include: isoniazid, rifamycins, pyrazinamide, ethambutol, streptomycin, levofloxacin, moxifloxacin, amikacin, capreomycin, kanamycin, cycloserine, ethionamide, para-aminosalicylate (PAS), clofazimine, bedaquiline, delamanid, linezolid, amoxicillin-clavulanate, and imipenem.☎	Required	L & P
	Positive interferon-gamma release assay (IGRA) test results (including, but not limited to: QuantiFERON®-TB Gold In-Tube, QuantiFERON® Plus, T-Spot.*TB*® test), for persons of any age; provide qualitative and quantitative positive IGRA results within 1 week of specimen collection.	--	L & P
Neisseria gonorrhoeae	Positive by any method for any specimen. If performed, antibiotic susceptibility test results should be reported, regardless of susceptibility pattern.	--	L & P
Neisseria meningitidis !	Positive culture or detected by nucleic acid amplification or positive immunohistochemistry or Gram-stain showing Gram-negative diplococci in CSF, blood, or any other sterile site or from petechial or purpuric lesion scrapings.	Required	L & P

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Pathogen [1]	Laboratory Tests and Results to Report to Public Health [2]	Send Isolate or Specimen [3]	Reporter [4]
Plasmodium species	Positive by any method for any specimen.	Required	L & P
Poliovirus ☎	Positive viral culture or detected by PCR for any specimen.	Required	L & P
Rabies virus: animal	The Tennessee Department of Health Laboratory conducts animal rabies testing statewide.	--	L & P
Rabies virus: human !	Testing is available through coordination with the Tennessee Department of Health Laboratory Services and CDC.	Required	L & P
Ricin toxin !	Positive by any method (including detection of DNA and presumptive identification of ricin toxin by fluoroimmunoassay) for any specimen.	Required	L & P
Rickettsia species (other than *R. typhus*)	• Detection of SFGR, including *R. rickettsii*, nucleic acid in a clinical specimen via amplification of a *Rickettsia* genus- or species-specific target by polymerase chain reaction (PCR) assays, • Elevated IgG antibody titer in one or more serology samples reactive with SFGR, including *R. rickettsii*, antigen by IFA*, • Demonstration of SFGR antigen in a biopsy or autopsy specimen by IHC, OR • Isolation of SFGR, including *R. rickettsii*, from a clinical specimen in cell culture and molecular confirmation (e.g., PCR or sequence). *Elevated IgM and positive ELISA/EIA for Rickettsia no longer require reporting	--	L & P
Rubella virus ☎	Positive IgM or rising IgG titer or detected by nucleic acid amplification or positive viral culture for any specimen.	Required	L & P
St. Louis encephalitis virus	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus- specific neutralizing antibodies. Any specimen.	--	L & P
Salmonella Typhi ☎	Positive by any method (including culture & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Salmonella species (other than *S.*Typhi)	Positive by any method (including culture & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Shigella species	Positive by any method (including culture & PCR, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Staphylococcus aureus, enterotoxin B-producing !	Positive by any method for any specimen.	Required	L & P
Staphylococcus aureus, methicillin-resistant **eip**	Isolation from a clinical specimen from a sterile site AND "non- susceptible" isolate identified (i.e., intermediate- or high-level resistance to cefoxitin, methicillin, nafcillin, or oxacillin) OR detection by nucleic acid amplification from a sterile site. Report only for Davidson County residents for EIP program; all NHSN reporting facilities (hospitals) will do statewide reporting to NHSN. Please include susceptibility test results and specimen source.	--	L & P
Staphylococcus aureus, Toxin-producing (TSST-1)	Positive by any method for any specimen.	--	L & P
Staphylococcus aureus, vancomycin non-susceptible : All forms ☎	Isolation from any clinical specimen AND "non-susceptible" isolate identified (i.e., intermediate- or high-level resistance to vancomycin). Please include all susceptibility test results with the numeric minimum inhibitory concentration values. Contact hai.health@tn.gov for clarification/questions.	Required	L & P
Streptococcus agalactiae	Positive culture or nucleic acid amplification from a normally sterile site.	--	L & P

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Streptococcus pneumoniae	Positive culture from any sterile site. Please include susceptibility test results with the numeric minimum inhibitory concentration values.	Required	L & P
Streptococcus pyogenes: Invasive Disease ☎, Toxin-producing	Positive culture or nucleic acid amplification from a normally sterile site, wound or muscle. Isolates from wounds will only be considered for Group A Streptococcal Invasive Disease when accompanied by necrotizing fasciitis (NF) or streptococcal toxic shock syndrome (STSS). Isolates from muscle will only be considered for Group A Streptococcal Invasive Disease.	Required	L & P
Treponema pallidum: Congenital ☎, Other	Report any treponemal or nontreponemal results, whether qualitative or quantitative, which are positive or reactive by any method. Report negative, non-reactive, or quantitative results for any testing associated with positive/reactive results. All reported non-treponemal results must include a titer value using standard notation (e.g., end-point reactivity at a serum dilution of 1:8 is reported as a titer of 8). All reactive non-treponemal screens should be confirmed with a standard treponemal test unless the patient had a known documented prior syphilis infection. Reports of reactive non-treponemal screens must also include either current treponemal test results (positive or negative) or prior confirmation information.	--	L & P
Trypanosoma cruzi	Positive by any method for any specimen.	--	L & P
Variola virus (orthopox virus) !	Positive by any method or suspected for any specimen.	Required	L & P
Vibrio cholerae (Toxigenic O1 or O139)	Positive by any method (including culture, PCR, & cholera toxin test, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Vibrio species (Non-toxigenic O1 or O139), *Grimontia hollisae, Photobacterium damselae*	Positive by any method (including culture, PCR, & cholera toxin test, **and excluding antibody positive tests**) for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Viral Hemorrhagic Fever viruses ! : Bunyaviruses, Crimean-Congo, Ebola, Guanarito, Junin, Lassa, Lujo, Machupo, Marburg, Sabia	Positive by any method for any specimen.	Required	L & P
West Nile virus	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus- specific neutralizing antibodies. Any specimen.	--	L & P
Yellow fever virus ☎	Positive IgM. Quantitative IgG indicating a positive test result. Isolation of virus or demonstration of specific viral antigen or nucleic acid. Virus- specific neutralizing antibodies. Any specimen.	Required	L & P
Yersinia pestis ☎	Positive by any method for any specimen. Include speciation results if known. Submit specimen/isolate to the Tennessee Department of Health Laboratory within 2 weeks of detection/isolation.	Required	L & P
Yersinia species (other than *Yersinia pestis*)	Positive by any method (including culture and PCR) for any specimen. Include speciation results if known.	Requested	L & P
Zika virus ☎	Report positive results by any method for any specimen. Submit specimens for positive IgM tests directly to CDC for further testing. No submission is requested or required for the Tennessee Department of Health Laboratory.	Required to CDC	L & P

Footnotes:

1 Timeframe for reporting: ! = phone immediately or ☎ = phone next business day | ^{eip} = report in 30 days via PH-1600 online or fax to HAI Emerging Infections Program (EIP) at (615) 741-3857. Contact hai.health@tn.gov for questions/clarification.

2 For most reportable diseases, a suspected/known case is reportable when the pathogen is isolated or detected from any specimen source (unless where otherwise indicated). A normally "sterile site" is defined as: blood, CSF, pleural fluid (includes chest fluid, thoracentesis fluid), peritoneal fluid (includes abdominal fluid, ascites), pericardial fluid, bone (includes bone marrow), joint (includes synovial fluid; fluid, needle aspirate or culture of any specific joint: knee, ankle, elbow, hip, wrist), internal body sites (specimen obtained from surgery or aspirate from one of the following: lymph node, brain, heart, liver, spleen, vitreous fluid, kidney, pancreas, or ovary). Screening cultures (e.g., nasal swabs, rectal, peri-rectal swabs) are included under "all isolates."

3 It shall be the responsibility of the director of a medical laboratory to submit isolates/specimens of designated microorganisms for confirmation, typing and/or antibiotic sensitivity. All specimens/isolates shall be accompanied by the following information: (a) Patient's full name, address, age, and sex. (b) Physician's name and address. (c) Anatomic source of culture. Refer to the Tennessee Department of Health Laboratory Services Directory of Services website for specimens needed for testing at https://www.tn.gov/health/health-program-areas/lab/directory-of-services.html.

4 The type of reporter responsible for reporting: L=Laboratory and P=Healthcare provider.